Raji/Human HVEM Stable Cell Line Data Sheet

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Catalog No.	Size
SCRAJ-STF108	2 × (1 vial contains ~5×10^6 cells)

• *Description*

The Raji/Human HVEM Stable Cell Line was engineered to express the full length human HVEM (Uniprot: Q92956-1), used to mimic cancer target cells. Surface expression of human HVEM was confirmed by flow cytometry.

• *Application*

- Useful for cell-based HVEM binding assay
- Useful as HVEM-expressing target cells in reporter gene assay

• *Cell Line Profile*

Cell line	Raji/Human HVEM Stable Cell Line
Host Cell	Raji
Property	Suspension
Complete Growth Medium	RPMI-1640 + 10% FBS
Selection Marker	NA
Incubation	37℃ with 5% CO_2
Doubling Time	16-20 hours
Transduction Technique	Lentivirus

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• *Materials Required for Cell Culture*

- RPMI Medium 1640 (ATCC, Cat. No. 30-2001)
- Fetal bovine serum (Gibco, Cat. No. 10091-148)
- Penicillin-Streptomycin (Gibco, Cat. No. 15140-122)
- Complete Growth Medium: RPMI-1640 + 10% FBS, 1%P/S
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, Cat. No. 430641)
- Cryogenic storage vials (SARSTEDT, Cat. No. 72.379.007)
- Thermostat water bath
- Centrifuge (Cence, Model: L550)
- Cell counter (MONWEI, Model: SmartCell200A Plus)
- CO2 Incubator (Thermo, Model: 3111)
- Biological Safety Cabinet (Thermo, Model: 1389)

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• *Recovery*

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize the risk of contamination, ensure the cap remains out of the water. Thawing should be completed quickly, typically within 3-5 minutes.
2. After thawing, promptly remove the vial from the water bath and decontaminate it by spraying with 70% ethanol. From this point onward, all operations must be performed under strict aseptic conditions.
3. Transfer the contents of the vial to a centrifuge tube containing 4.0 mL of complete growth medium.
4. Count viable cells and centrifuge at approximately 1000 rpm for 5 minutes.
5. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh complete growth medium. Adjust the cell density of the suspension to 1×10^6 viable cells/mL and transfer cells to an appropriate size vessel.
6. Incubate at 37°C with 5% CO_2 incubator.

• *Subculture*

Cell viability may be low after thawing, and full recovery (viability >90%) may take up to 1-2 weeks. Once the cell density reaches approximately 1.5×10^6 viable cells/mL, adjust the density to a range of 1×10^5-2×10^5 viable cells/mL by either adding the fresh culture medium or replacing the existing complete growth medium. Avoid allowing the cell density to exceed 2×10^6 cells/mL, as this may negatively impact cell performance in subsequent passages. T-75 flasks are recommended for subculturing.

• **Subculturing Frequency:** It is recommended to subculture every 3-4 days, adjusting the frequency based on the cell density in your specific culture system.

Note: After recovery, maintain the cells for 1-2 passages in the complete growth medium not containing the selection marker, if the cells are in good condition (viability >90%), transition to the culture medium containing the selection marker during subculturing.

• *Cryopreservation*

1. Count viable cells and harvest the cell suspension.
2. Centrifuge at 1000 rpm for 5 min at room temperature and resuspend cells in ice cold freezing medium to a concentration of 5×10^6 to 1×10^7 cells/mL.
3. Aliquot the cell suspension into cryogenic storage vials. Place the vials in a programmable cooler or an insulated box placed in a –80°C freezer overnight, then transfer to liquid nitrogen storage for long-term storage.

Note: It is recommended to establish a cell bank at the earliest possible passage for long-term use.

• *Storage*

Cells must be received in a frozen state on dry ice and should be transferred to liquid nitrogen or a –80°C freezer immediately upon receipt. If stored in a –80°C freezer, it is recommended to limit the storage period to no more than two weeks. For long-term preservation, transfer the cells to liquid nitrogen is highly recommended.

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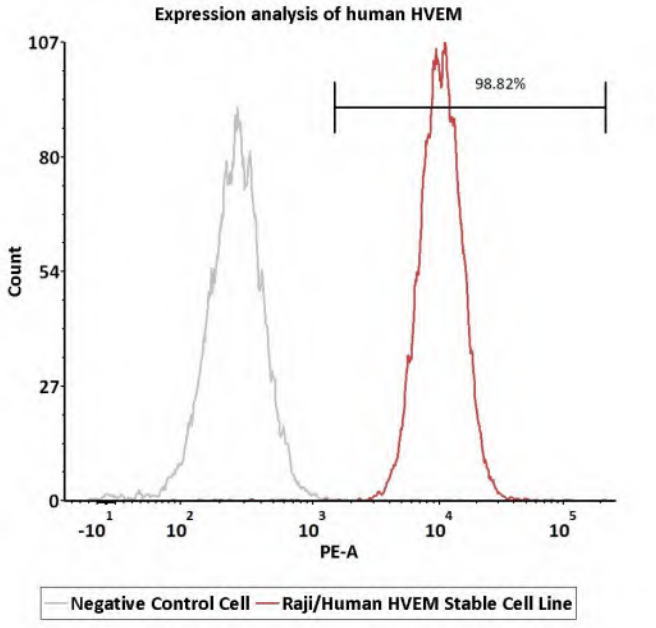
- ***Receptor Assay***



Catalog No.	Stable Cell Line	MFI for HVEM (PE)
NA	Negative Control Cell	254.53
SCRAJ-STF108	Raji/Human HVEM Stable Cell Line	9907.80

Fig1. Expression analysis of human HVEM on Raji/Human HVEM Stable Cell Line by FACS. Raji/Human HVEM Stable Cell Line or negative control cell were stained with PE-labeled anti-Human HVEM antibody.

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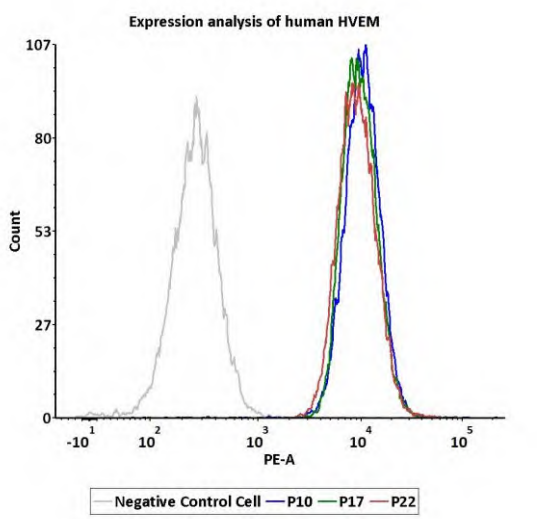
- ***Passage Stability***



Passage	MFI for HVEM (PE)
P10	9927.77
P17	9131.60
P22	8637.84

Fig2. Passage stability analysis of receptor expression by FACS. Flow cytometry surface staining of human HVEM on Raji/Human HVEM Stable Cell Line demonstrates consistent mean fluorescent intensity across passage 10-22.

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• *Related Products*

Products	Cat.No.
HEK293/Human PD-L1, GFP Tag Stable Cell Line	CHEK-ATP002
HEK293/Human 4-1BB Ligand / TNFSF9 Stable Cell Line	CHEK-ATP039
HEK293/Human 4-1BB / TNFRSF9 Stable Cell Line	CHEK-ATP038
Human PD-1/LAG-3 (Luc) Jurkat Reporter	SCJUR-STF063
Human PD-1 (Luc) Jurkat Reporter Cell	SCJUR-STF064
Human LAG-3 (Luc) Jurkat Reporter Cell	SCJUR-STF065
Raji/Human PD-L1 Stable Cell Line	SCRAJ-STT075
Raji/Human CD155 Stable Cell Line	SCRAJ-STT076
CHO/Human LILRB4 Stable Cell Line	SCCHO-ATP087
HEK293/Human LILRB4 Stable Cell Line	CHEK-ATP088
CHO/Human LIGHT Stable Cell Line	SCCHO-ATP109
CHO/Human BTLA Stable Cell Line	SCCHO-ATP110
HEK293/Human PD-1 Stable Cell Line	CHEK-ATP143
HEK293/Human HVEM Stable Cell Line	CHEK-ATP147
HEK293/Human NKp46 Stable Cell Line	CHEK-ATP153
HEK293/Human ITPRIPL1 Stable Cell Line	CHEK-ATP203
Human NKp46 (Luc) Jurkat Reporter Cell	SCJUR-STF130