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Raji/Human CD19 Knockout Stable Cell Line

| Catalog No. | Size |
|--------------|--|
| SCRAJ-STT216 | $2 \times (1 \text{ vial contains } \sim 5 \times 10^{6} \text{ cells})$ |

• Description

The Raji/Human CD19 Knockout Stable Cell Line was generated from Raji cells by CRISPR/Cas9-mediated knockout of human CD19 (Gene ID:930). The expression level of human CD19 was confirmed by flow cytometry. Mutated sequences of human CD19 produced by non-homologous end joining (NHEJ) were confirmed through genomic sequencing.

• Application

• Useful for cell-based CD19 target-specific analysis

• Cell Line Profile

| Cell line | Raji/Human CD19 Knockout Stable Cell Line | |
|------------------------|---|--|
| Host Cell | Raji | |
| Property | suspension | |
| Complete Growth Medium | RPMI Medium 1640 + 10% FBS | |
| Selection Marker | NA | |
| Incubation | 37°C with 5% CO ₂ | |
| Doubling Time | 18-22 hours | |
| Transduction Technique | Lentivirus | |



• Materials Required for Cell Culture

- PRMI-1640 Medium (ATCC, Cat. No. 30-2001TM)
- Fetal bovine serum (Gibco, Cat. No. A5669701)
- Penicillin-Streptomycin (Gibco, Cat. No. 15140-122)
- Phosphate Buffered Saline (1X) (HyClone, Cat. No. SH30256.01)
- Culture Medium: RPMI Medium 1640 + 10% FBS, 1% P/S
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, Cat. No. 430641)
- Cryogenic storage vials (SARSTEDT, Cat. No. 72.379.007)
- Thermostat water bath
- Centrifuge (Cence, Model: L550)
- Cell counter (MONWEI, Model: SmartCell200A Plus)
- CO₂ Incubator (Thermo, Model: 3111)
- Biological Safety Cabinet (Thermo, Model: 1389)

• Recovery

- 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize the risk of contamination, ensure the cap remains out of the water. Thawing should be completed quickly, typically within 3-5 minutes.
- 2. After thawing, promptly remove the vial from the water bath and decontaminate it by spraying with 70% ethanol. From this point onward, all operations must be performed under strict aseptic conditions.
- 3. Transfer the contents of the vial to a centrifuge tube containing 4.0 mL of culture medium.
- 4. Count viable cells and centrifuge at approximately 1000 rpm for 5 minutes.
- 5. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh culture medium. Adjust the cell density of the suspension to 1×10^6 viable cells/mL and transfer cells to an appropriate size vessel.
- 6. Incubate at 37° C with 5% CO₂ incubator.



• Subculture

Cell viability may be low after thawing, and full recovery (viability >90%) may take up to 1-2 weeks. Once the cell density reaches approximately 1.5×10^6 viable cells/mL, adjust the density to a range of 1×10^5 - 2×10^5 viable cells/mL by either adding the fresh culture medium or replacing the existing culture medium. Avoid allowing the cell density to exceed 2×10^6 cells/mL, as this may negatively impact cell performance in subsequent passages. T-75 flasks are recommended for subculturing.

• **Subculturing Frequency:** It is recommended to subculture every 3-4 days, adjusting the frequency based on the cell density in your specific culture system.

• Cryopreservation

- 1. Count viable cells and harvest the cell suspension.
- 2. Centrifuge at 1000 rpm for 5 min at room temperature and resuspend cells in ice cold freezing medium to a concentration of 5×10^6 to 1×10^7 cells/mL.
- 3. Aliquot the cell suspension into cryogenic storage vials. Place the vials in a programmable cooler or an insulated box placed in a -80°C freezer overnight, then transfer to liquid nitrogen storage for long-term storage.

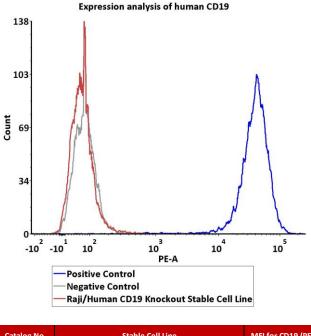
Note: It is recommended to establish a cell bank at the earliest possible passage for long-term use.

• Storage

Cells must be received in a frozen state on dry ice and should be transferred to liquid nitrogen or a -80° C freezer immediately upon receipt. If stored in a -80° C freezer, it is recommended to limit the storage period to no more than two weeks. For long-term preservation, transfer the cells to liquid nitrogen is highly recommended.



• Receptor Assay



| Catalog No. | Stable Cell Line | WHI TOP CD19 (PE) |
|--------------|---|-------------------|
| NA | Negative Control Cell | 74.34 |
| NA | Positive Control Cell | 39262.23 |
| SCRAJ-STT216 | Raji/Human CD19 Knockout Stable Cell Line | 63.92 |

Fig1. Expression analysis of human CD19 on Raji/Human CD19 Knockout Stable Cell Line by FACS.

Cell surface staining was performed on Raji/Human CD19 Knockout Stable Cell Line using PE-labeled antihuman CD19 antibody. The Raji cells were stained with PE-labeled anti-human CD19 antibody as the positive control cell. The Raji cells were stained with PE-labeled isotype control antibody as the negative control cell.



• Sequencing Analysis

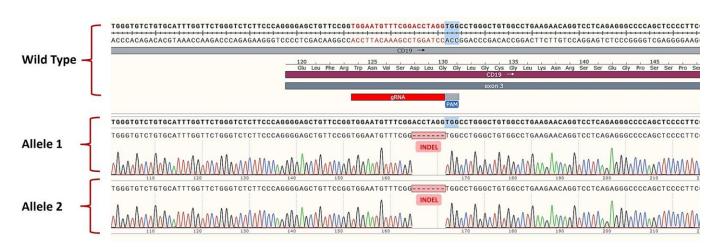


Fig2. Genomic Sequencing of human CD19 in the Raji/Human CD19 Knockout Stable Cell Line.

Sanger sequencing was used for analysis of CRISPR-mediated mutations. The sequencing results demonstrated that the selected sgRNA worked effectively with Cas9 on human CD19 gene in the Raji/Human CD19 Knockout Stable Cell Line.



• Related Products

| Products | <u>Cat.No.</u> |
|---|----------------|
| HEK293/hClaudin-18.2 Cell Line | CHEK-ATP033 |
| HEK293/hGPRC5D Cell Line | CHEK-STP042 |
| HEK293/Human TROP-2 Stable Cell Line | CHEK-ATP036 |
| HEK293/Human Nectin-4 Stable Cell Line | CHEK-ATP035 |
| HEK293/Human Anti-CD19 Stable Cell Line | CHEK-ATS056 |
| CHO/Human GPRC5D Stable Cell Line | CCHO-STP078 |
| HEK293/Human CEACAM5 Stable Cell Line | CHEK-ATP083 |
| HEK293/Human ROR1 Stable Cell Line | CHEK-ATP084 |
| HEK293/Human Transferrin R Stable Cell Line | CHEK-ATP089 |
| HEK293/Human DLL3 Stable Cell Line | CHEK-ATP090 |
| HEK293/Human FOLR1 Stable Cell Line | CHEK-ATP091 |
| HEK293/Human Glypican-3 (GPC3) Stable Cell Line | CHEK-ATP092 |
| CHO/Human DLL3 Stable Cell Line | SCCHO-ATP111 |
| CHO/Human Glypican-3 (GPC3) Stable Cell Line | SCCHO-ATP112 |
| HEK293/Human Transferrin Stable Cell Line | CHEK-ATP115 |
| HEK293/Human NAPI-IIb Stable Cell Line | CHEK-ATP116 |
| HEK293/Human Mesothelin Stable Cell Line | CHEK-ATP119 |
| CHO/Human Mesothelin Stable Cell Line | SCCHO-ATP120 |
| CHO/Human STEAP1 Stable Cell Line | SCCHO-ATP121 |
| HEK293/Human ENPP3 Stable Cell Line | CHEK-ATP122 |
| HEK293/Human LRRC15 Stable Cell Line | CHEK-ATP123 |
| HEK293/Human Claudin-1 Stable Cell Line | CHEK-ATP124 |
| HEK293/Human Integrin alpha V beta 6 Stable Cell Line | CHEK-ATP125 |
| HEK293/Human B7-H4 Stable Cell Line | CHEK-ATP126 |
| HEK293/Human Cadherin-6 Stable Cell Line | CHEK-ATP127 |
| HEK293/Human LY6G6D Stable Cell Line | CHEK-ATP137 |
| HEK293/Human Claudin-6 Stable Cell Line | CHEK-ATP138 |
| HEK293/Human Claudin-9 Stable Cell Line | CHEK-ATP139 |



• Related Products

Products

CHO/Human c-MET Stable Cell Line HEK293/Human c-MET Stable Cell Line HEK293/Human EGF R Stable Cell Line HEK293/Human ErbB3 Stable Cell Line HEK293/Human ErbB2 Stable Cell Line HEK293/Human uPAR Stable Cell Line CHO/Human uPAR Stable Cell Line HEK293/Human CD19 Stable Cell Line HEK293/Human STEAP1 Stable Cell Line CHO/Human B7-H3 (4Ig) Stable Cell Line CHO/Human CD79A&CD79B Stable Cell Line CHO/Human CD79B Stable Cell Line HEK293/Human Cadherin-17 Stable Cell Line HEK293/Human EpCAM Stable Cell Line HEK293/Human TPBG Stable Cell Line CHO/Cynomolgus Glypican-3 (GPC3) Stable Cell Line HEK293/Human GUCY2C Stable Cell Line HEK293/Human SEZ6 Stable Cell Line HEK293/Human FAP Stable Cell Line HEK293/Human PSMA Stable Cell Line HEK293/Human PTK7 Stable Cell Line HEK293/Human MCAM Stable Cell Line HEK293/Human GPC3 ∆HS Stable Cell Line HEK293/Human c-MET&ErbB3 Stable Cell Line HEK293/Human BCMA Stable Cell Line

<u>Cat.No.</u>

SCCHO-ATP141 CHEK-ATP146 CHEK-ATP148 CHEK-ATP149 CHEK-ATP150 CHEK-ATP151 SCCHO-ATP152 CHEK-ATP003 CHEK-ATP154 SCCHO-ATP169 SCCHO-ATP170 SCCHO-ATP171 CHEK-ATP173 CHEK-ATP175 CHEK-ATP176 SCCHO-ATP179 CHEK-ATP182 CHEK-ATP183 CHEK-ATP184 CHEK-ATP185 CHEK-ATP186 CHEK-ATP195 CHEK-ATP212 CHEK-ATP217 CHEK-ATP218